Investigation of subclinical mastitis cases in GCSAR Damascus goats from Humeimeh research station

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ABSTRACT

The study was conducted to evaluate the performance of California mastitis test, electrical conductivity measurement and lactose content as markers for subclinical-intramammary infection (IMI) in goats. It was performed at Humeimeh research station for Damascus goats, General commission for scientific agricultural research (GCSAR) from February to March 2011 and done on dairy goats aged 2 to 6 years. Milk samples were taken from 134 dairy goats during morning milking according to habitual procedure and screened for evidence of subclinical mastitis by California Mastitis Test (CMT), electrical conductivity measurement (EC) and lactose content (LC) of milk. Positive (+1 to +3) CMT and/or high electrical conductivity milk samples were subjected to bacteriological examination to distinguish between healthy (absence of mastitis agents) and infected (presence of contagious or environmental mastitis agents). The overall prevalence of subclinical mastitis was found to be 24.6%. Percentage of positive (+1 to +3), suspected and negative CMT reactions were 76.85, 8.95 and 14.2%, respectively. The values of electrical conductivity for negative, suspected and positive (+1, +2 and +3) CMT milk samples were 3.93 ± 0.64, 4.47 ± 0.61, 4.68 ± 0.72, 4.81 ± 0.76 and 6.56 ± 0.85 ms/cm, respectively. T-test has been shown statistical differences in California mastitis test reactions and electrical conductivity readings, and not statistically for milk lactose content, between positive and negative results for bacterial growth. Intramammary infection was found in 40.9% milk samples caused by Staphylococcus (75%), alone E. coli (16.6%) or associated with streptococci (8.3%). Correlation coefficient between CMT reactions with electrical conductivity readings was R = 0.494. Higher coefficient correlation for electrical conductivity measurement and Kappa agreement index (R = 0.491 and 0.42 respectively) with positive bacterial growth milk samples than for CMT (R = 0.35 and 0.12 respectively) make the electrical conductivity measurement more compatible with the results of bacteriological analysis than CMT despite a slight increase of CMT sensitivity, and reduced number of real negative cases.

Keywords: Damascus goats, subclinical mastitis, California CMT test, electrical conductivity, lactose content of milk.

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INTRODUCTION

Mastitis in small ruminants has a major effect in reducing both yield and quality of milk leading to strong economic losses (Stuhr and Aurlich, 2010). Moreover, mastitis is one the most frequently cited reason for culling (Tangorra et al., 2010). Many nations have taken attention to quality of the milk in small ruminants, especially after using the genetic improvement programs (Gabina and Barillet, 1991) and after intensifying of production systems for these animals (Haenlein, 1993). Attention is not longer based on the essential compounds of milk, but on the other indices providing a clear picture of the quality of the produced milk and the health of the udder such as the number of somatic cells and the level of electrical conductivity and others. The causes of mastitis are many...
and varied; it is usually caused by injury but also due to incorrect feeding, unsanitary, and poor living conditions. Damage is caused to the udder by improper milking practices. Genetic and physiological factors (such as race, age, estrus, season of milking, milk yield or stage of lactation), increase mastitis susceptibility. Others as health standards and requirements increase somatic cells and electrical conductivity in the milk of goats under non-infectious conditions (Stuhr and Aurlich, 2010).

It seems that the most common methods (Somatic cell count, California Mastitis Test and electrical conductivity) for the control of udder health and udder infections are of limited value for goats [5]. Contrary to strong correlation between indirect diagnostic tests: CMT results and somatic cells count, and the results of the bacterial milk culture in ewes there is a problem in the interpretation of cell count and the results of California mastitis test of in goats. California Mastitis Test (CMT) is a common indirect method of measuring Somatic cell count (SCC) in cows, but some authors claim that CMT is an unreliable method for diagnosing intramammary infection (IMI) in goats (Schaeren and Maurer, 2006; Bergonier et al., 2003). Other studies, however, report that CMT may be useful for detection of healthy udders (Karzis et al., 2007; Petzer et al., 2008). The electrical conductivity is considered an rapid and non-direct indicator of the health of the udder in goats (Romero et al., 2010). It refers to the changes in permeability due to injury of the udder, associated with rising concentrations of ions and cations, as sodium Na⁺ And chloride Cl⁻ secreted in the milk of infected udder halves (Krömer, 2007). Because of changes in the permeability of the udder tissues and its fluctuations of readings, electrical conductivity is not usually used in goat's milk. Electrical conductivity levels (Norberg, 2004) are affected by a variety of factors, the most important is the temperature of the milk samples (Alonso and Mimor, 2006), clinical subclinical mastitis (Hamann and Zecconi, 1998; Norberg, 2004), microbial contamination of milk (Daumoras and Kyrs, 2006), the acidity of the milk (Lanzanova et al., 1993; Mucchetti et al., 1994), animal race (Park, 1991), milking season (Dinsmor et al., 1998; Regin et al., 2002), the somatic cells content of the milk (Mucchetti et al., 1994; Fah et al., 2001; Bansal et al., 2007). The value of electrical conductivity of milk changes during stage of milk production within the same lactation season (Sheldrake et al., 1983). The type of the devices used to measure of electrical conductivity also plays very important role in the results (Maatje et al., 1983; Onyango et al., 1988; Crame, 1987).

There are reported values of electrical conductivity got an average values of electrical conductivity 6.6 ± 0.5 ms/cm (Schuppel and Schwope, 1999) close to values 6.33 ± 0.75 ms/cm obtained by Boulaba (2009). According to Tangorra et al. (2010) there is no absolute threshold reliable permitting to distinguish between the inflamed and non-inflamed udder glands. Also Ryffel et al. (2007) could not put comparable limits values in goat milk. The fact that the direct count of somatic cells in milk samples helped to diagnose of subclinical mastitis and therefore the percentage of infection in milk samples taken from herds has been increased from 20-50% to 70.2% (Boscos et al., 1996). The levels of lactose content in milk decrease after the udder becoming infected and partially permeable for lactose formed in the udder from glucose and galactose (Stuhr and Aurlich, 2010). Noticing also that Jendretzke (2009) indicated that the lactose level in milk is depending of some factors like estrus, feeding, lactation number and age. It ranges between 3.8 and 4.6% and falls at the end of milking season.

Milk samples can be considered infected if they accompanied by a positive ≥+1 CMT reaction, white cell count ≥ 0.63 million cells / ml and the level of lactose ≤ 4.33%, or chloride content ≥ 0.109% (Upadhyaya and Rao, 1993).

In this study, indirect diagnose subclinical mastitis methods are used: measurement of somatic cell count (SCC) by California Mastitis Test (CMT), electrical conductivity (EC) measurements lactose content LC analysis in milk is evaluated. The purpose of the present study was to compare the performance of CMT, EC and LC as possible markers for subclinical-intramammary infection (IMI) in goats, to assess the prevalence of subclinical-intramammary infection in Humeimeh Research Station for Damascus goats and moreover to see how well indirect measurements of SCC (CMT and EC) correspond to growth of pathogens in the udder.

MATERIALS AND METHODS

The study was conducted at Humeimeh Research Station for Damascus goats/General Commission for Agricultural Scientific Research (GCSAR). Animals that were used for the study were born in the station but their origin was from Damascus region. Animals are usually herded twice a day (mornings before getting out and after being herded afternoon). The does were milked twice per day (during morning and evening hours). The lactating does were given barley grain, wheat bran and cotton seed cake and concentrate during milking period. The study was conducted from February to march 2011. A total of 134 foremilk samples (about 10 ml) were collected from dairy goat aged 2 to 6 years once during morning milking from one half or two halves according to habitual procedure and screened for evidence of subclinical mastitis by California Mastitis Test (CMT), electrical conductivity measurement and lactose content of milk. Positive (+1 to +3) CMT and/or high electrical conductivity milk samples were subjected to bacteriological examination to distinguish between healthy (absence of mastitis agents) and infected (presence of contagious or environmental mastitis agents). Milk samples for the bacterial culture of (+1 to +3) CMT and/or highly electrical conductivity reading were taken after cleaning , washing and disinfecting the udder with alcohol 70.0% in sterile plastic bottles excluding the first fore-milk stream.

The following indirect methods for the detection of subclinical mastitis have been used:

Electrical conductivity measurements

This was conducted using Suntex SC-17 which measures electrical
Table 1. Grading CMT reactions or defining CMT scores.

<table>
<thead>
<tr>
<th>CMT Reaction</th>
<th>Symbol</th>
<th>Interaction</th>
<th>Cell count/ml</th>
</tr>
</thead>
<tbody>
<tr>
<td>unchanged</td>
<td>0</td>
<td>Negative</td>
<td>68,000</td>
</tr>
<tr>
<td>Precipitate a mild viscous form disappears with continuous movement</td>
<td>T</td>
<td>Effect</td>
<td>268,000</td>
</tr>
<tr>
<td>distinct precipitated 'mucus' Disappears with continuous movement</td>
<td>+1</td>
<td>Weak positive</td>
<td>800,000</td>
</tr>
<tr>
<td>directly thickness of the mixture with the emergence of a gel</td>
<td>+2</td>
<td>Clear</td>
<td>2,560,000</td>
</tr>
<tr>
<td>Forming gelatinous with convex surface</td>
<td>+3</td>
<td>Strong</td>
<td>≥ 10,000,000</td>
</tr>
</tbody>
</table>


conductivity or current flow resistance of milk in ohms( mho) or Siemens (S) per meter (ms/m = 10^6 mmho/cm), and that between the negative and positive sensitive poles for measuring the temperature of samples automatically.

California CMT

The main advantages with CMT are that it is quick, cheap and simple and that it is an "animal-side". This test was performed as described (Schalm et al., 1971; Ikram, 1997) to classify the milk samples in intact and infected udders before performing bacterial growth culture (Tangorra et al., 2010). The principle of the test is the reactive reagent (BOVI.VET CMT-Test from KRUSESE Danish Company) which is a solvent reacts with the DNA of somatic cell nuclei after the dissolution of their outer wall and the nucleus cell wall with the formation of filamentous mass which is proportional with the somatic cells count. The presence of neutrophils, which are directly related to glandular irritation leads to increase of somatic cell count (Sheldrake et al., 1983). The presence of positive interactions does not necessarily indicate the presence of mastitis. The bacterial toxins that reach the bloodstream leads also to massive destruction of white blood cells without being associated with a positive result in CMT (Wagner et al., 2007).

To interpret the results of CMT test (Shearer and Harris, 1992) (Table 1): a) Trace or weak result of CMT (viscous to mucus disappears with continuous movement) indicate suspicion of the udder inflammation (≤268 × 10^4 cell/ml), which means that the udder has just recovered from a previous infection or that there is an evidence of inflammation of the udder. In both cases the test must repeated to verify the presence of inflammation or make another diagnostic test. b) Clearly positive results without any doubt (directly thickness of the mixture with the emergence of a gel): indicate evalutative inflammatory response associated with advanced high somatic cells count (≥800 × 10^4 cell/ml). This may be due to mechanical injury of the mammary gland or more likely caused by germ infection. Causes led to inflammation of the udder (still electricity or problems related to machine milking must be defined and corrected immediately.

Lactose content of milk

This was measured once in positive and negative in positive and negative milk samples for microbial growth by Melchoscan S 50 From Foss Company.

Bacterial analysis

CMT positive (+1 to+3) and/or high electrical conductivity milk samples, which numbered 66 samples were cultured once at the

Table 2. Goat's status according to the results of bacterial growth as a diagnostic test.

<table>
<thead>
<tr>
<th>CMT</th>
<th>bacterial growth</th>
<th>Status</th>
</tr>
</thead>
<tbody>
<tr>
<td>≥ +1 reaction</td>
<td>Positive result</td>
<td>Real positive</td>
</tr>
<tr>
<td>≥ +1 reaction</td>
<td>Negative result</td>
<td>False positive</td>
</tr>
<tr>
<td>negative reaction</td>
<td>Positive result</td>
<td>False negative</td>
</tr>
<tr>
<td>negative reaction</td>
<td>Negative result</td>
<td>real negative</td>
</tr>
</tbody>
</table>

Laboratory of the Animal Health Department, Aleppo Directorate of Agriculture on McConkie culture and blooded agar plates which were incubated at 37°C for 16 to 24 h, and re-evaluated at 48 h on milk samples, the bacteria were identified by means of typical colony morphology according to the standard modalities of Barth (2009).

If moderate to high growth of a major udder pathogen was found the sample would be diagnosed as positive for bacterial analysis.

Prevalence of the subclinical, accuracy, sensitivity and specificity values, positive and negative predictive and positive likelihood ratio for the CMT and EC

Prevalence of the subclinical, accuracy, sensitivity and specificity values positive and negative predictive and Positive likelihood ratio for the CMT and EC were calculated (McGee, 2002; Reid et al., 1998) after goats have been classified into groups according to CMT reactions and electrical conductivity readings (threshold of electrical conductivity is ≥ 4.75 ms/cm) depending on the result of bacterial growth as a diagnostic test (Table 2).

Prevalence

The prevalence is the proportion of true (a) positive (positive for either diagnostic test or bacterial culture), and false(c) negative (positive diagnostic test but negative bacterial culture) in the population or:

\[
\frac{a + c}{a + b + c + d} \times 100
\]

Accuracy

The proportion of true positive (a) and true negative (d) in the population or:

\[
\frac{a + d}{a + b + c + d} \times 100
\]
Sensitivity

The proportion of true positive (a) in true positive (a) and false negative (c) or:

\[ \frac{a}{a + c} \times 100 \]

Specificity

The proportion of true negative in true negative and false positive or:

\[ \frac{d}{b + d} \times 100 \]

Positive predictive value (PPV)

The proportion of true positive in true positive and false positive or:

\[ \frac{a}{a + b} \times 100 \]

Negative predictive value (NPV)

The proportion of true negative in true negative and false negative or:

\[ \frac{d}{c + d} \times 100 \]

Positive likelihood ratio (LR+)

\[ \text{Sensitivity} \div (1 - \text{Sensitivity}) \]

Negative likelihood ratio (LR-)

\[ \frac{(1 - \text{Sensitivity}) \div \text{Specificity}}{\frac{c}{d} \div \frac{a + c}{b + d}} \]

Pre-test probability

The proportion of true positive and false negative in the total number of cases:

\[ \frac{a + c}{a + b + c + d} \times 100 \]

Pre-test odds

\[ \frac{\text{prevalence}}{(1 - \text{prevalence})} \text{ or } \frac{\text{pre-test probability}}{(1 - \text{pre-test probability})} \]

Post-test probability

\[ \frac{\text{post-test odds}}{(\text{post-test odds} + 1)} \]

Post-test odds

Pre-test odds \times \text{Likelihood ratio}

Positive posttest probability

\[ \frac{a}{a + b} \]

Negative posttest probability

\[ \frac{c}{c + d} \]

Statistical analysis

T-test was carried out using Genstat 7.2 (Genstat Seventh Edition, 2004) to study the differences in CMT reactions and electrical conductivity readings between positive and negative bacterial growth results. T-test for independent for CMT group reactions was also performed to test the differences between the means of electrical conductivity readings using the program (StatPac, 2011).

Agreement measure (Cohen kappa index)

The performance of the modern technologies or test' evaluation need to be interpreted whether the differences between different tests due to the methods used or for the opinions public commentators. The Cohen kappa agreement index is preferred statistically because it measures the agreement between the results of diagnostic methods, and interprets what's real or accounts for chance (Bland and Altman, 1986; Kundel and Polansky, 2003) (Table 3).

Cohen's kappa coefficient was used to measure the agreement between in hand: CMT, electrical conductivity EC measurements with the results of the bacteriological analysis in the other hand (<0 no agreement, 0 to 0.2 slight agreement, 0.2 to 0.4 fair agreement, 0.4 to 0.6 moderate agreement, 0.6 to 0.8 substantial agreement, and 0.8 to 1 almost perfect agreement).

RESULTS

Statement of assorted goats basing on the results of California mastitis test and electrical conductivity

Goats were assorted according to the results of CMT and electrical conductivity measurements basing on the result of bacterial growth as a diagnostic test and so it was obtained four groups symbolized A to D. As shown in Table 4, an increase of the truth negative cases for the
Table 3. Data entering model to measure the Cohen kappa index for the agreement.

<table>
<thead>
<tr>
<th>Diagnostic test positive</th>
<th>Yes</th>
<th>No</th>
<th>Total</th>
</tr>
</thead>
<tbody>
<tr>
<td>Negative diagnosis test</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Yes</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Positive bacterial growth, positive result of diagnostic test</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>No</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Positive bacterial growth but negative result of diagnostic test</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Total</td>
<td></td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

Table 4. Issue of goat’s assortment basing on the results of CMT and EC.

<table>
<thead>
<tr>
<th>Symbol</th>
<th>Statement</th>
<th>California test</th>
<th>Electrical conductivity</th>
</tr>
</thead>
<tbody>
<tr>
<td>A</td>
<td>Positive for the disease with a positive diagnostic test or true positive</td>
<td>30</td>
<td>28</td>
</tr>
<tr>
<td>B</td>
<td>Negative for the disease with positive diagnostic test or false positive</td>
<td>71</td>
<td>32</td>
</tr>
<tr>
<td>C</td>
<td>Positive for the disease with negative diagnostic test or false-negative</td>
<td>3</td>
<td>5</td>
</tr>
<tr>
<td>D</td>
<td>Negative for the disease, as well as diagnostic test or true negative</td>
<td>30</td>
<td>69</td>
</tr>
<tr>
<td>Total</td>
<td></td>
<td>134</td>
<td></td>
</tr>
</tbody>
</table>

Table 5. Values of electrical conductivity (ms/cm) for CMT reactions.

<table>
<thead>
<tr>
<th>Symbol</th>
<th>0</th>
<th>1</th>
<th>2</th>
<th>3</th>
</tr>
</thead>
<tbody>
<tr>
<td>SD + Mean</td>
<td>3.93 ± 0.64</td>
<td>4.47 ± 0.61</td>
<td>4.68 ± 0.72</td>
<td>4.81 ± 0.76</td>
</tr>
<tr>
<td>SE</td>
<td>0.15</td>
<td>0.16</td>
<td>0.092</td>
<td>0.13</td>
</tr>
</tbody>
</table>

measure of electrical conductivity is faced by an increase of false positive cases for CMT.

Prevalence of udder subclinical mastitis in GCSAR’ Damascus goat research stations

The prevalence of subclinical mastitis in GCSAR’ Damascus goat research station (for CMT and EC) which is the proportion of true positive and false in the population according to (Byrt et al., 1993 to Feinstein and Cicchetti, 1990) was amounted to 24.6%.

CMT results

The percentage of positive reactions (+1 to +3) and suspected and negative CMT was 76.85, 8.95 and 14.2%, respectively.

Results of measuring the electrical conductivity according to CMT reactions

The values of electrical conductivity for negative, suspected and positive (+1, +2 and +3) CMT milk samples were 3.93 ± 0.64, 4.47 ± 0.61, 4.68 ± 0.72, 4.81 ± 0.76 and 6.56 ± 0.85 ms/cm, respectively. The t-test showed highly significant differences (P ≤ 0.002) in CMT reactions and in electrical conductivity readings between positive and negative bacterial growth milk samples. T-test for independent groups showed using (Genstat Seventh Edition, 2004) also significant differences (P ≤ 0.05) between means of electrical conductivity values (Table 5) for CMT inter-group reactions between negative and +1 CMT (P ≤ 0.05), between negative and +2 and +3 CMT (P ≤ 0.01), between suspected and +3 CMT (P ≤ 0.002), and between +1 and +3 CMT (P ≤ 0.002), while no differences were observed between means and standard deviations for electrical conductivity readings between suspected and +3 CMT inter-groups and for inter-group CMT and between suspected and +1 and +2 CMT inter-groups, and also between +2 and +1 CMT inter-groups.

Correlation coefficients between CMT reactions with electrical conductivity readings, and between both electrical conductivity readings and CMT reactions with positive bacterial growth in milk samples were (R = 0.494, R = 0.491 and R = 0.35 respectively). T-test has been shown statistical differences in California mastitis test reactions and electrical conductivity reading between positive and negative results for bacterial growth.

Lactose content of milk

Low levels of lactose content were recorded in positive milk samples for bacterial growth compared to negative
Table 6. Statistically performance of CMT and EC.

<table>
<thead>
<tr>
<th>Type of diagnostic test</th>
<th>CMT</th>
<th>EC</th>
</tr>
</thead>
<tbody>
<tr>
<td>Accuracy %</td>
<td>44.8</td>
<td>72.4</td>
</tr>
<tr>
<td>Sensitivity %</td>
<td>90.9</td>
<td>84.8</td>
</tr>
<tr>
<td>Specificity %</td>
<td>29.7</td>
<td>68.3</td>
</tr>
<tr>
<td>Positive predictive value PPV %</td>
<td>29.7</td>
<td>46.7</td>
</tr>
<tr>
<td>Negative predictive value NPV %</td>
<td>30.9</td>
<td>93.2</td>
</tr>
<tr>
<td>Positive likelihood ratio LR +</td>
<td>1.29</td>
<td>2.68</td>
</tr>
<tr>
<td>Negative likelihood ratio LR -</td>
<td>0.31</td>
<td>0.22</td>
</tr>
<tr>
<td>Pre-test probability;</td>
<td>0.246</td>
<td></td>
</tr>
<tr>
<td>Post- Probability</td>
<td>0.297 or 3/10</td>
<td>0.467 or 5/10</td>
</tr>
<tr>
<td>Pre-test odds</td>
<td>0.327</td>
<td></td>
</tr>
<tr>
<td>Post-test odds</td>
<td>0.42</td>
<td>0.88</td>
</tr>
<tr>
<td>Cohen's kappa index</td>
<td>0.1217</td>
<td>0.4168</td>
</tr>
<tr>
<td>Pearson correlation factor with the result of bacterial analysis</td>
<td>0.35</td>
<td>0.49</td>
</tr>
</tbody>
</table>

Table 7. Cohen's kappa index for CMT and EC according to Sharma et al. (2010).

<table>
<thead>
<tr>
<th>Measurement of the electrical conductivity</th>
<th>Positive</th>
<th>Total</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Yes</td>
<td>No</td>
</tr>
<tr>
<td>Positive</td>
<td>28</td>
<td>32</td>
</tr>
<tr>
<td>Negative</td>
<td>5</td>
<td>69</td>
</tr>
<tr>
<td>Total</td>
<td>33</td>
<td>101</td>
</tr>
</tbody>
</table>

<table>
<thead>
<tr>
<th>California test</th>
<th>Positive</th>
<th>Total</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Yes</td>
<td>No</td>
</tr>
<tr>
<td>Negative</td>
<td>30</td>
<td>71</td>
</tr>
<tr>
<td>No</td>
<td>3</td>
<td>30</td>
</tr>
<tr>
<td>Total</td>
<td>33</td>
<td>100</td>
</tr>
</tbody>
</table>

Results of the bacteriological analysis in suspected and positive milk samples

Intramammary infection, defined as growth of udder pathogens, was found in 40.9% milk samples caused by contagious pathogens represented by Staphylococcus staphylococci (75%). In addition to environmental pathogens represented by alone E. coli (16.6%) or associated with streptococci (8.3%) all sensitive to the following antibiotics: Cephalosporin, Cephalexin, Ciprofloxacin, Ofloxacin, Enrofloxacin, Doxycycline Sellin - neomycin, Amikacin (cannot be administrated to pregnant does).

Statistical measures of the performance of CMT and EC

Table 6 shows high accuracy and specificity of the measure of the electrical conductivity despite of high CMT sensitivity. Noting also that an increase of truth negative cases for electrical conductivity was faced by similar increase of false positive cases of CMT (Table 6). However, Cohen's kappa index which was obtained for the measuring electrical conductivity is closer to 1 than for CMT (slight) and therefore it was more compatible (Sharma et al., 2010) (Table 7).

DISCUSSION

The prevalence of subclinical mastitis in GCSAR' Damascus Goat Research Station Humeimeh was amounted to 24.6%. The overall prevalence of subclinical mastitis in goats at Tigray Regional State, North Ethiopia was found to be 28.14% (Gebrewahid et al., 2012).

The percentage of positive reactions (+1 to +3) and suspected and negative CMT were 76.85, 8.95 and 14.2%, respectively. The percentage of positive reactions was very close to the results obtained (76.7%) in dairy goats in the province of Morogoro in Tanzania by Mbilu (2007), but higher than what obtained (Upadhyaya and Rao, 1993) for (+1 to +3) CMT (67.7%) and therefore lower for suspected (trace) and negative CMT reactions (32.8%). Noticing that there are significant differences between CMT readings in milk samples as well as between udder halves whose somatic cell count is ≥0.750 × 10³ cells/ml, as well as between infected and uninfected udders as indicated (Petzer et al., 2008).

The values of electrical conductivity for negative, suspected and positive (+1, +2 and +3) CMT milk samples were 3.93 ± 0.64, 4.47 ± 0.61, 4.68 ± 0.72, 4.81...
The electrical conductivity readings for (+3) CMT was (6.56 ± 0.85 ms/cm) closed to the average level obtained by Deiad et al. (2010) for normal milk (6.33 ms/cm) for the yield milk of 1564.2 g. Fluctuations in electrical conductivity measurements for CMT ratings may be due to the type of device used for measurements and temperature of the samples (Alonso and Mimor, 2006; Maatje et al., 1983; Onyango et al., 1988; Crame, 1987). This is supported by the fact that it would be necessary of time-series analysis of historical data of the electrical conductivity among the latest highly recorded 20th readings during the beginning and middle of the end lactation season and the health status of the two udder halves and the comparison between the values of different halves through lactation season to obtain satisfactory results (Tangorra et al., 2010).

T-test for independent groups using (Genstat Seventh Edition, 2004) showed also significant differences (P ≤ 0.05) between means of electrical conductivity values for CMT inter-group reactions between negative and +1 CMT (P ≤ 0.05), between negative and +2 and +3 CMT (P ≤ 0.01), between suspected and +3 CMT (P ≤ 0.002), and between +1 and +3 CMT (P ≤ 0.002), while no differences were observed between means and standard deviations for electrical conductivity readings between suspected and +3 CMT inter- groups and for inter-group CMT and between suspected and +1 and +2 CMT inter-groups, and also between +2 and +1 CMT inter-groups.

As Kifaro et al. (2009) no significant influence on lactose content in positive milk samples for bacterial growth was observed (4.1 ± 0.18% vs. 4.15 ± 0.19%), contrary to Leitner et al. (2004) who reported significantly lower concentration of lactose in the infected glands (4.17 ± 0.13) in comparison to uninfected one (4.7 ± 0.1). The measurement in this study was done once and could not be repeated and therefore there were no statistical differences between positive or negative milk samples for bacterial growth.

Intramammary infection was found in 38.9% milk samples. There was found lower Intramammary infection IMI than in other publications (Persson and Olofsson, 2011), because only 18% of all udder halves had IMI, where the proportion of udder halves with subclinical IMI in goats ranged from 23 to 70% (Leitner et al., 2004; Persson and Olofsson, 2011; McDougall and Prosser, 2010). The lower proportion of Persson and Olofsson (2011) study might be the result of good udder health in Sweden. Staphylococcus (75%) isolated in this present study correspond to Schaefer and Maurer (2006) and Leitner et al. (2004) who indicated that Staphylococcus aureus is the most prevalent pathogens of the mammary gland in goats (60 to 80%) in goats. In addition to isolated Staphylococcus there were environmental pathogens represented by alone E. coli (16.6%) or in association with streptococci (8.3%). Noticing that Gebrewahid et al. (2012) had isolated from CMT positive samples: Staphylococcus aureus (27.7%), Escherichia coli (17.0%) and Streptococcus (10.63%) in goats at Tigray Regional State, North Ethiopia.

Correlation coefficients between CMT reactions with electrical conductivity readings, and between both electrical conductivity readings and CMT reactions with positive bacterial growth in milk samples were (R = 0.494, R =0.491 and R = 0.35 respectively). T-test has been shown statistical differences in California mastitis test reactions and electrical conductivity readings between positive and negative results for bacterial growth. According to Schaefer and Maurer (2006) in 20 to 30% of the cases the SCC is ≥750,000 cells/ml. Also, the relation between California Mastitis Test (CMT) reactions and udder infections is not very close. Moreover, over 20% of mammary halves infected with CN (coagulase-negative) Staphylococcus showed negative CMT reactions. On the other hand, 25% of samples from mammary halves without a proven infection reacted positively (Schaefer and Maurer, 2006). This confirms the idea what pointed it to (Koop et al., 2011) that the sensitivity of the bacterial analysis is very low compared to the count of somatic cells SCC. The prevalence of mastitis by S. aureus in dairy goats must be higher than what obtained by bacterial analysis.

The low number of false positive, relative high accuracy and positive PPV and negative NPV predictive values, relative high positive likelihood ratio LR+ and relative high agreement of Cohen’ kappa index, and Pearson correlation between measuring electrical conductivity with bacterial culture (32 vs. 71 for CMT, 72.4%, 46.7%, 93.2%, 0.4168, 2.68 and 0.49 respectively) make this test more compatible with the results of bacteriological analysis than CMT (Sharma et al., 2010; Martin et al., 1987) despite the high number of real negative of electrical conductivity in comparison to CMT (69 vs.30).

Conclusions

1. Both California test and electrical conductivity give nearly the same number of true positive and false-negative but different number of false positive and true negative.

2. The electrical conductivity measurement is more compatible with the results of bacteriological analysis than CMT.

3. It can be concluded that the CMT does not reflect really the health status of the udder; it is more influenced than EC by different factors cited by the authors like increased days in milking, parity and reduced milk production.

4. The fact that the positivity CMT does not mean necessarily the infection of the udder. This is supported by the fact that most of the milk samples were bacteriologically negative.

5. Lactose content in positive or negative milk samples
for bacterial growth was unchanged (4.1 ± 0.18 and 4.15 ± 0.19%, respectively). It needs probably repeated measurements.

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